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Individual free fatty acids have unique associations with inflammatory biomarkers, insulin resistance and insulin secretion in healthy and gestational diabetic pregnant women

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ABSTRACT

Objective We investigated the relationships of maternal circulating individual free fatty acids (FFA) with insulin resistance, insulin secretion and inflammatory biomarkers during mid-pregnancy.

Research design and methods The data were drawn from a prospective cohort of generally healthy pregnant women (n=1368, African-American 36%, Hispanic 48%, Caucasian 16%) in Camden, NJ. We quantitatively determined 11 FFAs, seven cytokine/adipokine, homeostatic model assessment of insulin resistance (HOMA-IR) and C-peptide levels from the fasting blood samples that were collected at 16 weeks of gestation. Multivariate analyses were performed along with separate analyses for each individual FFA.

Results High HOMA-IR ($p<0.001$) and C-peptide ($p<0.0001$) levels were positively associated with a twofold to fourfold increased risk for developing gestational diabetes mellitus (GDM). Negative relationships were found with specific FFAs (molecular percentage, palmitoleic, oleic, linolenic, myristic acids) and HOMA-IR and C-peptide levels ($p<0.01$ to $p<0.0001$). In contrast, palmitic, stearic, arachidonic, dihomo- γ -linolenic (DGLA) and docosahexaenoic acids were positively associated with HOMA-IR and C-peptide ($p<0.01$ to $p<0.0001$). The individual FFAs also predicted cytokine/adipokine levels. For example, women who had elevated DGLA (highest quartile) were twice as (adjusted OR 2.06, 95% CI 1.42 to 2.98) likely to have higher interleukin (IL)-8 ($p<0.0001$) levels. Conversely, women with high palmitoleic, oleic, and linolenic acid levels had reduced odds (≥ 2 -fold, $p<0.01$ to $p<0.001$) for having higher IL-8, IL-6 or tumor necrosis factor- α levels.

Conclusion Our results suggest that maternal individual FFAs uniquely affect insulin resistance and secretion. The effects are either direct or indirect via modulation of the inflammatory response. Modifying the composition of FFAs may help in reducing the risk of GDM.

INTRODUCTION

Resistance to the actions of insulin on glucose and fat metabolism has been observed in normal pregnancy, particularly during late gestation.^{1 2} Excessive insulin resistance

Significance of this study

What is already known about this subject?

- Previous studies were limited to the relation of total concentrations of free fatty acids (FFAs) with insulin resistance/secretion in pregnant women with and without gestational diabetes mellitus (GDM).
- How specific individual FFAs are involved in the regulation of glucose homeostasis and modulate inflammatory response in normal pregnancy and in women prior to the diagnosis of GDM remains uncertain.

What are the new findings?

- In this prospective and well characterized cohort of young pregnant women, we show that specific individual FFAs have strong, but opposite effects on insulin resistance and insulin secretion by mid-pregnancy.
- Different associations between specific FFAs and inflammatory biomarkers (cytokine/adipokine) were also observed.

How might these results change the focus of research or clinical practice?

- Our data suggest that individual FFAs have unique metabolic impacts in healthy pregnant women and in women prior to the diagnosis of GDM.
- The recognition and modification of circulating FFAs composition could be important for reducing the risk of GDM and other pregnancy complications related to subclinical inflammation and insulin resistance.

(IR) and significant β -cell dysfunction are key components for the development of gestational diabetes mellitus (GDM).^{3–5} How IR develops during pregnancy is not fully understood. Circulating free fatty acids (FFA) have been recognized as one of the most important factors that contribute to IR and also alter insulin secretion.^{1 6 7} Previous studies by Boden and Chen demonstrated that experimentally created acute elevations

in circulating total FFA concentration cause increases in IR in healthy non-pregnant subjects, and type 2 diabetes mellitus (T2DM) and in pregnant women with and without GDM.^{1 7 8} Elevated total FFA levels significantly increase insulin secretion in normal subjects, but not in T2DM and GDM; here insulin secretion is defective.^{1 6-8} Thus, the relationship of the individual FFAs, instead of total FFA, to IR and insulin secretion in human pregnancy needs further investigation.

Strong evidence indicates that chronic low-grade inflammation which is characterized by increased circulating cytokine levels is another key factor associated with IR.^{4 9} FFAs have emerged as an important link for explaining inflammatory-related conditions.¹⁰⁻¹² Increased total FFA levels induced whole body proinflammatory responses and impaired endothelial function in non-pregnant healthy subjects.^{9 13} Supplementation with n-3 fatty acids (FA) in obese pregnant women resulted in decreased cytokine levels in both placenta and adipose tissues.¹⁴ Although in vitro studies have reported that individual FAs modulate the inflammatory response in cultured cells,^{15 16} results are equivocal. Whether maternal individual FFAs affect inflammatory cytokine levels differently in human pregnancy is not known.

The purpose of the present study was to examine the relationships of individual FFAs with measures of IR (homeostatic model assessment of insulin resistance, HOMA-IR), insulin secretion indicated by C-peptide level, and inflammatory biomarkers from the Camden Study, a well-characterized prospective cohort of generally healthy pregnant women at high risk of adverse maternal-fetal outcomes. We hypothesized that individual FFAs have (1) different relationships with IR and insulin secretion and (2) different associations with inflammatory cytokine/adipokine levels in human pregnancy.

RESEARCH DESIGN AND METHODS

Study design and population

The data were collected as part of the Camden Study, a prospective cohort study of maternal nutrition and pregnancy outcome in young, generally healthy women residing in one of the poorest cities in the continental United States.^{17 18} The study was conducted in the Osborn Family Health Center at Our Lady of Lourdes Hospital in Camden, NJ. Informed written consent was obtained from each participant after explanation of the nature and purpose of the study.

Study participants were enrolled between October 1996 and June 2006 for prenatal care. A total of 3.5% of the women who had serious non-obstetric problems (eg, lupus, type 1 or 2 diabetes, seizure disorders, malignancies, acute or chronic liver disease, drug or alcohol abuse and psychiatric problems) were found not to be eligible at screening. Eighty percent of the patients who were eligible agreed to participate in this study. Of the 1703 pregnant women who had all measurements assayed, we also excluded participants who were diagnosed with

pre-eclampsia (n=186) or delivered a preterm infant (n=149) since both conditions are potentially associated with metabolic disturbance and/or increased inflammation.¹⁹⁻²¹ Twenty-five women who were diagnosed with GDM and also had complication of pre-eclampsia (n=13) or preterm delivery (n=12) were not excluded. A final group of 1368 women were included in this analysis.

Data collection

Data of socioeconomic, demographic and lifestyle characteristics were obtained by interview at entry to care (13.4±5.2 weeks of gestation, mean±SD), and updated at 20 and 28 weeks of gestation. Ethnicity was self-defined. Gestational duration was assessed with an algorithm based on a participant's last normal menstrual period confirmed or modified by early ultrasound. Body mass index (BMI) was computed based on self-reported pregravid weight and measured height at entry to prenatal care (kg/m²).

Diagnosis of GDM and definition of normal controls

The diagnosis of GDM was made by a two-step approach. Patients were initially screened by a 50 g oral glucose challenge test at the 24–28 weeks of gestation. A 100 g oral glucose tolerance test (OGTT) was performed on that subset of women exceeding the glucose threshold value (>140 mg/dL at 1 hour). The diagnostic criteria for GDM were based on the Carpenter/Cooustan conversion as recommended by the American Diabetes Association.²² Participants with a normal glucose screen test were defined as controls.

Sample collection and analytical procedures

Fasting blood samples (>8 hours) collected at entry to care (15.8±5.2 weeks of gestation) were refrigerated and centrifuged at 4°C. The serum and plasma samples were aliquoted and stored at -70°C until assayed.

Measurement of FFAs

We measured the plasma concentration and relative abundance of 11 FFAs (myristic, palmitic, stearic, palmitoleic, oleic, linoleic, α -linolenic, dihomo- γ -linolenic (DGLA), arachidonic, eicosapentaenoic (EPA) and docosahexaenoic (DHA) acids). Total lipids were extracted by a modified Folch method and analyzed by gas chromatography/mass spectrometry (Agilent Technologies 5973 MS/6890 GC, Santa Clara, CA) as described previously.²³ An internal standard heptadecanoic acid (C17:0) was added to each sample before the extraction of FFAs to identify the FFA fraction from other lipid classes which was separated by thin layer chromatography. A nitroterephthalic acid-modified polyethylene glycol capillary column (ZB-FFAP, Phenomenex, Torrance, CA) was used. Peak retention times were identified by injecting known standards (Sigma-Aldrich, Louis, MO). The precision and reproducibility was monitored by the analysis of reinjection on the same samples and the samples from different batches.

The peak area of FFA was identified as the percentage of total area under the peaks and absolute concentration

of FFA was quantified by multiplying each individual FFA in relative value (%) by the total FFA concentration ($\mu\text{mol/L}$) determined by an enzymatic assay kit (Wako Chemicals USA, Richmond, VA).

Assays of cytokine and adipokine

Cytokines (interleukin (IL)-6, IL-8, IL-10, tumor necrosis factor alpha (TNF- α) and granulocyte-macrophage colony-stimulating factor) and adipokines (adiponectin and resistin) were analyzed by two panels using the Luminex xMAP technology (Luminex, Austin, TX) on the MagPix system. Serum cytokine concentrations were determined by the human high sensitivity T-cell 5-plex magnetic bead panel and adipokines were determined by the human adipokine 2-plex magnetic bead panel (EMD Millipore, Billerica, MA). According to the manufacturing protocol, there was no or negligible cross-reactivity between the antibodies for an analyte and any of the other analytes in each of the panels. The average intracoefficient variation (CV) was 5.5% (range 4.6%–7.0%) and the average inter-CV was 11.2% (range 8.2%–13.2%) for cytokines. The intra-CV was 4.4% and 5.5%, the inter-CV was 10.1% and 11.3% for adiponectin and resistin, respectively. All analyses were performed according to the manufactures' protocols and were done in duplicate. The concentrations were calculated from best fit standard curves generated from calibrators for each analyte in each assay by Milliplex Analyst V.5.1.

Parameters of glucose and insulin homeostasis

Fasting plasma glucose was measured with the glucose oxidase method (Sigma Diagnostics, St Louis, MO) on a Multiscan analyzer (BioTek, Winooski, VT). Serum insulin was determined by radioimmunoassay (RIA) using a kit with a specific antibody that cross-reacts only minimally (<0.2%) with proinsulin and has a high sensitivity (2 mU/mL or 12 pmol/L). Plasma C-peptide was determined by an RIA kit with high sensitivity (0.1 ng/mL or 0.033 nmol/L) and low cross activity to proinsulin (<4%) (Linco, St Charles, MO). The intra-CV and inter-CV were 3.3% and 6.1% for C-peptide, 4.9% and 7.2% for insulin, 3.3% and 5.3% for glucose. We used C-peptide concentration as an indicator of insulin secretion because it is cosecreted on an equimolar basis from the β -cell with insulin and is not extracted or metabolized by the liver.^{3 18} HOMA-IR was computed as index of IR.^{24 25}

Dietary data

A 24 hours' recall of the previous day's diet was obtained at entry to care, week 20 and 28 gestation processed with databases from the Campbell Institute of Research and Technology (Campbell Soup Company) in Camden as described previously.²³ The database generates data for more than 70 nutrients and 19 FAs using the US Department of Agriculture Nutrient Database for Standard Reference and the Continuing Survey of Food Intakes by Individuals. The mean of three 24-hour recalls was used

to calculate the total daily intake of energy, carbohydrate, protein, total fat, individual FAs, saturated FA (SFA), monounsaturated FA and polyunsaturated FA (PUFA) accordingly.

Statistical analysis

Parametric statistics were calculated for the continuous variables, and χ^2 tests for independence were used for the categorical variables. Log10 transformations were performed if the data were positively and highly skewed. Multiple linear regression analyses were used to examine the relationships of individual FFAs (expressed as molecular percentage, %) with the continuous outcome variables (HOMA-IR and C-peptide level). Separate regression models were employed to estimate the relationships of each of the independent variables with the outcome variables. The same types of analyses were also performed with the women who were diagnosed with GDM during later gestation and in normal controls (online supplementary tables 1 and 2). Multiple logistic regressions were performed to estimate the ORs and 95% confidence intervals (CI) for the elevated levels of the glucose metabolism parameters (\geq median level) with GDM at entry (prior to the clinical diagnosis of GDM).

We also conducted multiple polytomous logistic regression analyses to estimate the associations of the elevated individual FFAs with the cytokine/adipokine levels. This is an extension of traditional logistic regression in multiple levels of an outcome variable that are all adjusted so that the ORs are estimated in the same model. The frequency distributions of the FFAs (%) were divided into quartiles, and logistic regression was used to estimate the OR of high (top quartile) FFAs with cytokine/adipokine levels in each quartile using the lowest quartile as the reference category. The linear trends of specific FFAs across the quartiles of cytokine/adipokine were also tested.

In order to examine the relation between dietary and circulating FFAs, we correlated (Pearson's r) plasma individual FFA (molecular %) with the corresponding dietary FA intake (as % of total fat intake) adjusted for energy intake.

All multivariable analyses were also conducted controlling for potential confounding variables which included maternal BMI, age, parity, cigarette smoking, and ethnicity. All of the levels of significance were two sided, and $p < 0.05$ was considered to be the lowest level of acceptable statistical significance. All of the statistical analyses were performed with SAS V.9.4 (SAS Institute).

RESULTS

The characteristics of the women at entry (~16 weeks' gestation) are shown in table 1. As expected, women who developed GDM during late gestation were older and had higher pre-pregnant BMI ($p < 0.0001$). There were more Hispanic and fewer African-American women in the GDM group ($p < 0.001$). The mean gestational age at blood sampling, parity and cigarette smoking were not

Table 1 Characteristics of study participants

Characteristic	All participants	GDM	Normal controls
n	1368	81	1287
Age (year)	22.08±5.24	26.01±6.05	21.90±5.03*
Pre-pregnant BMI (kg/m ²)	25.73±6.22	30.08±6.42	25.37±6.21*
Obesity	250 (18.27)	35 (43.21)	215 (17.92)*
Gestational age at blood sampling (weeks)	15.79±5.24	16.74±5.19	15.72±5.21
Nulliparas	516 (37.72)	29 (35.80)	487 (37.83)
Ethnicity			
Hispanic	656 (47.66)	47 (58.02)	609 (47.37)
African-American	490 (35.81)	14 (17.28)	476 (36.98)
Caucasian and other	222 (16.22)	20 (24.69)	202 (15.67)†
Cigarette smoking	260 (19.00)	17 (20.99)	243 (18.89)
Plasma glucose (mmol/L)	4.39±0.02	4.98±0.09	4.34±0.02*
Serum insulin (pmol/L)	129.73±2.43	161.82±9.79	127.79±2.78†
Plasma C-peptide (nmol/L)	0.74±0.02	1.20±0.06	0.71±0.02*
HOMA-IR	3.75±0.08	4.99±0.32	3.67±0.08*

Data are mean±SD for continuous variables and n (%) for categorical variables.

Fasting plasma glucose, insulin, C-peptide and HOMA-IR (means±SE) were adjusted for maternal age, pre-pregnant BMI, and gestational age at blood sampling.

*P<0.0001 versus GDM.

†P<0.001 versus GDM.

.BMI, body mass index; GDM, gestational diabetes mellitus; HOMA-IR, homeostatic model assessment of insulin resistance.

significantly different between the GDM women and normal controls ($p>0.05$ for each). The concentrations of fasting plasma glucose, C-peptide, serum insulin and HOMA-IR were significantly higher in the GDM women than in the normal controls ($p<0.001$ to $p<0.0001$ for each).

Further analysis by multiple logistic regressions showed that elevated (\geq median entry level) glucose (≥ 4.38 mmol/L) (adjusted OR (AOR) 2.15, 95% CI 1.30 to 3.56), insulin (≥ 99.73 pmol/L, AOR 2.49, 95% CI 1.41 to 4.40), C-peptide (≥ 0.54 nmol/L, AOR 4.43, 95% CI 2.25 to 8.72) and HOMA-IR (≥ 2.75 , AOR 2.71, 95% CI 1.51 to 4.86) were significantly associated with increased risk for the subsequent development of GDM ($p<0.01$ to $p<0.0001$ for each).

Individual FFAs with HOMA-IR and C-peptide

The HOMA-IR and C-peptide levels were regressed on the linear combination of the potential confounding variables (age, pre-pregnant BMI, cigarette smoking, ethnicity and parity) in multiple regression analyses. The estimated regression coefficients for the multiple models are presented in table 2. There were significant negative relationships of several individual FFAs with HOMA-IR (β coefficient was -0.666 , -0.100 , -1.317 and -0.309 for 1% increases in palmitoleic or oleic or α -linolenic or myristic acids, respectively, $p<0.01$ to $p<0.0001$ for each). Inverse relations were also found with respect to C-peptide (β coefficient was -0.457 , -0.069 , -0.984 and -0.211 ng/mL C-peptide for 1% increases in palmitoleic or oleic or α -linolenic or myristic acids, respectively, $p<0.001$ to $p<0.0001$ for each).

In contrast, significant positive associations of several FFAs with HOMA-IR were observed (β coefficient was 0.065, 0.084, 0.778 and 0.269 for palmitic, stearic, DGLA, and arachidonic acids, respectively, $p<0.001$ to $p<0.0001$ for each); similar relationships were found with C-peptide ($p<0.0001$ for each). There was no significant association of linoleic acid and EPA with HOMA-IR. EPA and DHA were both positively related to C-peptide ($p<0.001$ and $p<0.0001$).

Separate analyses were performed for women who subsequently developed GDM and the normal controls. The results from the normal controls were identical to those for all participants (online supplementary table 1). Similar results were also found for the GDM women, but the magnitude of these relationships was not statistically significant (online supplementary table 2). The lack of significance most likely was probably attributable to the small sample size of GDM group.

The trends of relationships (positive and negative) between individual FFAs and HOMA-IR or C-peptide in non-GDM women who complicated with pre-eclampsia and preterm delivery are similar to those shown with GDM and non-GDM normal controls, but significant association was only found in palmitoleic acid ($p<0.01$) in women with preterm delivery; the association was significant in palmitoleic, oleic, α -linolenic and stearic acids in women with pre-eclampsia ($p<0.05$ to $p<0.001$, data not shown).

Individual FFAs and inflammatory biomarkers

A summary for the FFAs, which were consistently showing linear associations with at least two cytokines/adipokines, is presented in table 3. Elevated individual FFAs (the highest quartile vs other quartiles pooled) were associated with inflammatory biomarkers in opposite directions (p for trend <0.05 to <0.0001 for each). Significant negative relationships were found for myristic, palmitoleic, linolenic and oleic acids. For example, women with higher (highest quartile) palmitoleic acid levels relative to the other quartiles had a 2.7-fold significant decrease for having a high IL-8 level (highest quartile vs lowest quartile, 15.1% vs 31.4%, AOR was 0.36, 95% CI 0.24 to

Table 2 Association of individual FFAs with HOMA-IR and C-peptide concentration

Independent variable	Molecular % (mean±SD)	Dependent variable			
		HOMA-IR β±SE	P value	C-peptide (ng/mL) β±SE	P value
Palmitoleic acid (16:1)	2.58±1.22	-0.666±0.070	<0.001	-0.457±0.044	<0.0001
Oleic acid (18:1)	26.37±7.42	-0.100±0.011	<0.0001	-0.069±0.007	<0.0001
Linoleic acid (18:2)	15.53±4.94	-0.031±0.017	NS	-0.022±0.011	<0.05
α-Linolenic acid (18:3)	0.60±0.27	-1.317±0.318	<0.0001	-0.984±0.200	<0.0001
Myristic acid (14:0)	1.91±0.85	-0.309±0.101	<0.01	-0.211±0.064	<0.001
Palmitic acid (16:0)	34.33±7.94	0.065±0.010	<0.0001	0.040±0.007	<0.0001
Stearic acid (18:0)	14.45±5.32	0.084±0.016	<0.001	0.059±0.010	<0.0001
DGLA (20:3)	1.20±0.75	0.778±0.108	<0.0001	0.677±0.067	<0.0001
Arachidonic acid (20:4)	2.02±1.22	0.269±0.069	<0.0001	0.229±0.043	<0.0001
EPA (20:5)	0.15±0.31	0.290±0.266	NS	0.557±0.168	<0.001
DHA (22:6)	0.89±0.55	0.303±0.152	<0.05	0.410±0.095	<0.0001

Models were adjusted for maternal age, pre-pregnant body mass index (BMI), parity, ethnicity and cigarette smoking. Separate model was performed for each independent variable with HOMA-IR or C-peptide level.

DGLA, dihomo- γ -linolenic acid; DHA, docosahexaenoic acid; EPA, eicosapentaenoic acid; FFA, free fatty acid; HOMA-IR, homeostatic model assessment of insulin resistance; NS, not significant.

0.55, p for trend <0.0001). For myristic acid, the AOR was 0.37 (95% CI 0.25 to 0.54). Similar results were found for the same group of FFAs with IL-6, TNF- α or resistin. Opposite relationships were found with respect to DGLA, palmitic acid and EPA (table 3). For example, having a high DGLA predicted a twofold increase in the odds of having a high IL-8 (AOR 2.06, 95% CI 1.42 to 2.98, p for trend <0.0001). However, no significant relationships of stearic, linoleic, and DHA with cytokine/adipokine levels were observed. Arachidonic acid was only associated with one marker of cytokine/adipokine, TNF- α (AOR 1.72, 95% CI 1.20 to 2.46), which is not listed in table 3.

For the FFAs that had significant linear relationships with cytokine/adipokine levels, we compared the effect of higher FFAs on each of the quartile of cytokine/adipokine levels (using the lowest quartile as reference) (figure 1). For the negative associations of linolenic and myristic acids, there was a trend for decreasing AORs across the IL-6, IL-8 and TNF- α quartiles. For the positive associations of DGLA and palmitic acid, the AORs increased across the quartiles for IL-8, TNF- α , and resistin.

We did not observe any association of FFAs (in molecular percentage) to adiponectin. Adiponectin was inversely correlated with maternal pre-pregnant BMI (Pearson's r =-0.283, p <0.0001), HOMA-IR (β coefficient -0.616, p <0.001) and C-peptide (β coefficient -0.567, p <0.0001). In addition, the adiponectin levels were negatively correlated with the absolute concentrations (μ mol/L) of palmitic acid (p <0.0001), stearic acids (p <0.01), DGLA (p <0.05) and total FFA (p <0.01). Compared with the lowest quartile, the highest quartile of palmitic acid was associated with decreasing levels of adiponectin (AOR 0.66, 95% CI 0.46 to 0.94) when BMI

was not controlled for. This finding was not significant after BMI was controlled for.

Individual FFAs and dietary fat intake

We correlated plasma individual FFA (molecular %) with the corresponding dietary FA intake (as % of total fat intake) adjusted for energy intake. Plasma myristic acid was minimally correlated with dietary intakes of myristic acid (r =0.100, p <0.0001), total fat (r =-0.07, p <0.05) and total PUFAs (r =-0.116, p <0.001). The correlations with other FFAs were non-significant and negligible.

CONCLUSIONS

In this prospective cohort of young pregnant women, we have shown different associations of circulating individual FFAs with measurements of IR and secretion. Most importantly, opposite relationships between specific FFAs and inflammatory biomarkers (cytokine/adipokine) were also observed. The results indicate that individual FFAs have unique metabolic impacts by mid-pregnancy.

Markers identifying GDM risk

There has been a lack of consistency in the identification of markers that predict GDM.^{26–28} We found that higher HOMA-IR or C-peptide or insulin or glucose concentration determined at ~16 weeks' gestation was associated with 2.7-fold to 4.4-fold increased risk for developing GDM independent of several traditional risk factors, including maternal age and BMI. Recently, in a secondary analysis of case-control study, Hinkle *et al* reported that elevated HbA_{1c} levels at 8–13 weeks' gestation were associated with GDM.²⁶ Another study reported that HOMA-IR at week 7–12 gestation was associated with GDM.²⁷ Thus, impaired glucose homeostasis seems to occur before

Table 3 Association of elevated individual FFAs (molecular percentage, %) with inflammatory biomarkers

High FFA and cytokine/adipokine*	Quartile 1 n=338	Quartile 2 n=349	Quartile 3 n=343	Quartile 4 n=338	AOR (95% CI)†	P for trend
Myristic acid (≥ 2.28 vs < 2.28)						
IL-6	30.9	27.5	21.6	20.1	0.58 (0.40 to 0.83)	<0.0001
IL-8	35.8	27.1	20.6	16.5	0.37 (0.25 to 0.54)	<0.0001
TNF- α	33.4	23.8	26.9	15.9	0.40 (0.27 to 0.59)	<0.0001
Resistin	19.3	27.5	26.5	26.2	1.30 (0.89 to 1.88)	NS
Palmitoleic acid (≥ 3.09 vs < 3.09)						
IL-6	24.2	28.3	27.1	20.4	0.67 (0.46 to 0.98)	<0.05
IL-8	31.4	26.9	26.6	15.1	0.36 (0.24 to 0.55)	<0.0001
TNF- α	22.6	27.3	34.1	16.0	0.56 (0.38 to 0.84)	<0.05
Resistin	29.2	25.4	24.5	20.9	0.55 (0.38 to 0.80)	<0.001
α-Linolenic acid (≥ 0.71 vs < 0.71)						
IL-6	31.3	23.0	26.2	19.5	0.50 (0.35 to 0.72)	<0.001
IL-8	28.6	25.4	25.1	20.8	0.67 (0.47 to 0.96)	<0.05
TNF- α	27.8	25.7	27.8	18.8	0.62 (0.43 to 0.89)	<0.05
Resistin	23.9	27.1	24.1	25.0	0.91 (0.64 to 1.29)	NS
Oleic acid (≥ 30.58 vs < 30.58)						
IL-6	24.1	24.9	29.8	21.2	0.75 (0.52 to 1.09)	NS
IL-8	28.7	27.5	26.9	16.8	0.50 (0.34 to 0.73)	<0.001
TNF- α	25.8	25.5	30.1	18.6	0.62 (0.43 to 0.90)	<0.05
Resistin	26.7	28.6	26.4	18.6	0.52 (0.36 to 0.76)	<0.001
DGLA (≥ 1.73 vs < 1.73)						
IL-6	23.5	25.5	25.8	25.2	1.08 (0.76 to 1.55)	NS
IL-8	17.9	23.2	29.1	29.7	2.06 (1.42 to 2.98)	<0.0001
TNF- α	19.5	25.1	25.7	29.8	1.83 (1.28 to 2.63)	<0.01
Resistin	20.8	23.2	30.5	25.5	1.22 (0.85 to 1.76)	NS
Palmitic acid (≥ 37.56 vs < 37.56)						
IL-6	22.5	24.4	23.8	29.5	1.49 (1.05 to 2.12)	<0.05
IL-8	23.7	22.2	23.7	30.3	1.51 (1.07 to 2.15)	<0.05
TNF- α	25.5	22.4	23.6	28.5	1.22 (0.86 to 1.73)	NS
Resistin	17.3	24.1	27.1	31.6	2.20 (1.40 to 2.92)	<0.0001
EPA (≥ 0.16 vs < 0.16)						
IL-6	22.7	23.9	24.5	29.0	1.47 (1.03 to 2.11)	<0.05
IL-8	21.2	23.7	24.9	30.2	1.76 (1.22 to 2.54)	<0.01
TNF- α	23.4	25.9	24.9	25.9	1.26 (0.87 to 1.81)	NS
Resistin	18.7	26.6	27.8	26.9	1.34 (0.92 to 1.95)	NS

*High FFA was defined as the highest quartile of FFA.

†Models were adjusted for maternal age, pre-pregnant body mass index (BMI), parity, ethnicity and cigarette smoking. AOR was estimated for patients who were in the highest quartile of FFA versus other quartiles pooled, association with inflammatory biomarkers in the top quartile using the lowest quartile as reference.

AOR, adjusted OR; DGLA, dihomo- γ -linolenic acid; EPA, eicosapentaenoic acid; FFA, free fatty acid; IL, interleukin; NS, not significant; TNF- α , tumor necrosis factor alpha;

the third trimester; our findings confirm that simple measurements may be useful for earlier identification of some women at risk for GDM.

Individual FFAs are associated with IR and insulin secretion differently

A particularly interesting finding in the present study was that specific individual fasting FFAs have large, but opposite effects on IR and C-peptide by mid-pregnancy

(table 2). The strongest negative relationship was for the essential FA, α -linolenic acid, whereas DGLA, an endogenously produced n-6 FA, was one of the most positive correlates to HOMA-IR. Similar results were observed for the FFAs and C-peptide.

Previous studies were limited to the relation of total FFA concentration with IR/secretion. Elevation in plasma total FFAs by infusion of lipid and heparin in healthy

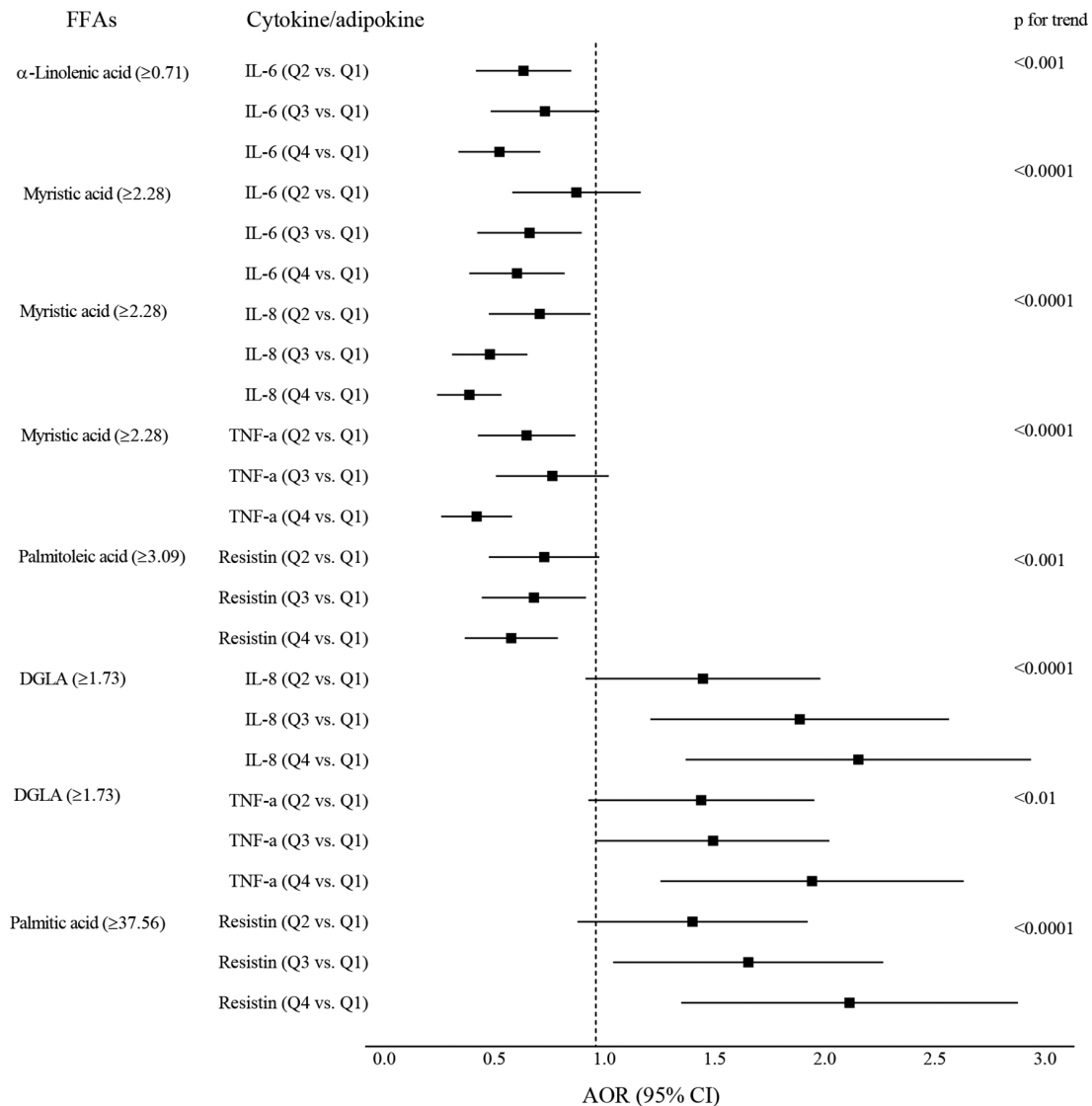


Figure 1 Linear associations of specific FFAs with inflammatory biomarkers. Square markers indicate adjusted ORs (AOR) which were estimated for patients who were in the highest quartile of FFA (vs other quartiles pooled), association with inflammatory biomarkers (comparing the quartiles 2–4 to the lowest quartile). Error bars indicate 95% CI. Models were adjusted for maternal age, pre-pregnant body mass index (BMI), parity, ethnicity and cigarette smoking. Separate models were performed for each FFA with each of the cytokines/adipokines. DGLA, dihomo- γ -linolenic acid; FFA, free fatty acid; IL, interleukin; TNF, tumor necrosis factor.

subjects, patients with type 2 diabetes, or pregnant women with and without GDM caused IR by reducing insulin-stimulated glucose uptake.^{6–9} Increased total FFA level also has a role as an insulin secretagogue in patients with diabetes and healthy individuals.^{5,6} Our findings support FFAs having significant relationships with IR and insulin secretion, and indicate that individual FFAs are differentially associated with IR and insulin secretion.

FAs have an important role in the regulation of insulin homeostasis. In a prospective cohort of American Indians, Lemaitre *et al* have shown that higher plasma level of ceramides or higher sphingomyelins (the sphingolipids species) with SFAs, such as palmitic acid, have different associations with fasting insulin, HOMA-IR or HOMA β -cell function.²⁵

We found that higher myristic acid, an SFA, was negatively correlated with having favorable relationships with both HOMA-IR and inflammation (tables 2 and 3). In non-pregnancy studies, serum myristic acid has positive or an unfavorable association with subclinical inflammation and with increased risk of T2DM.^{10,29} For unknown reasons, our results are contrary to previous observations in the literature. Additional studies are needed to understand the underlying mechanisms.

Individual FFAs are associated with inflammatory cytokine/adipokine differently

Another significant finding from the current study was that elevated individual FFAs were significantly associated with several major inflammatory markers, including IL-6, IL-8,

and TNF- α and resistin, in different directions (table 3). Inflammation is a widely known contributor linked to increased IR.^{9 11 12 15 30} However, little is known about the effects of individual FFAs as modulators of inflammation during pregnancy.

Many studies have reported significant effects for total FFA levels and/or n-3 FAs in inflammation.^{9 14 31} Acute increases in total FFA concentrations induce intracellular and total cellular inflammation in healthy subjects.⁹ Supplementation of n-3 FAs (EPA and DHA) in non-pregnancies reduces blood levels of inflammatory markers.³¹ However, in obese pregnant women, supplementation of DHA and EPA significantly reduces the inflammatory response in maternal placental and adipose tissues, but plasma inflammatory markers do not show changes, except with respect to a reduction of C-reactive protein concentration ($p < 0.05$).¹⁴ Thus, the effects of FFAs on the inflammation are not always detectable in blood levels of inflammatory markers.

Likewise in our study, not all individual FFAs were associated with inflammatory biomarkers. It is generally believed that the n-3 FAs have anti-inflammatory and SFAs have proinflammatory properties.^{11 16 31} However, a recent study suggested that differences in chain lengths of individual SFAs had an opposite effect on inflammation.¹⁰ Currently, there are insufficient data in human studies to predict of which specific FAs will affect inflammatory status and how the effect can be detected.^{9 15 30 32} We observed strong relationships, particularly with several FFAs representing mono-unsaturated, n3, n6, and saturated FFAs, suggesting the unique impacts of FFAs on chronic inflammation during pregnancy.

Finally, we did not find significant relationships between FFAs (in %) and adiponectin. The association of adiponectin to several SFA concentrations was dependent on maternal BMI. Although adiponectin has a strong anti-inflammatory property and the concentration is decreased in GDM,^{33 34} the published results on its relationship with FFAs are mixed. A report from Spain studied 17 FAs (without known lipid class sources), and only found that palmitic acid and myristic acid were negatively correlated with adiponectin in non-pregnant women ($p = 0.02$ and $p = 0.007$).³⁵ A significant inverse association between circulating adiponectin and plasma non-esterified FA or palmitic acid levels was found during an experimental elevation of intravascular triglyceride lipolysis in healthy men.³⁴ In contrast, acute lowering of total FFA concentration pharmaceutically was associated with a decrease in the adiponectin level.³⁶ Further studies are also needed to better understand the association of FFAs with adiponectin in pregnancies.

Strengths and limitations

Strengths of our study are a well-characterized large prospectively designed cohort including the measurements of indices of IR/secretion, inflammatory biomarkers as well as FFAs which have been validated and widely adopted in experimental and epidemiologic studies.^{3 8 9 24 25} There are some potential limitations. First, we included 25 women diagnosed with GDM and also complicated with either

pre-eclampsia or preterm delivery because GDM is the focus of this study. We could have performed a separate analysis for GDM with and without pre-eclampsia or preterm delivery if the sample size was larger. Second, circulating FFA composition has been considered as a biomarker of fat intake, but the reports on their relationship have been inconsistent.^{37–40}

We were unable to find strong relationships between plasma FFAs and dietary fat intake by 24 hours' recalls. Plasma FFA levels are the end products of both exogenous (dietary fat intake) and metabolism endogenously (lipolysis, lipogenesis and oxidation), it is not feasible for our epidemiologic study to determine and distinguish the kinetics of fat metabolism. In addition, the time differences between blood sample collection and the assessment of dietary intakes could also contribute to the results. Lastly, we have used diagnostic criteria for GDM from the American Diabetes Association.²² By using the new criteria from the hyperglycemia and adverse pregnancy outcome (HAPO) can result in a significantly increased number of women diagnosed with GDM.^{41 42} The participants in our cohort were enrolled prior to HAPO study, we did not obtain data of a 75 g OGTT for all participants, and therefore, we are unable to detect additional cases of GDM. In the current study, our main focus is to examine the association of individual plasma FFAs with the cardiometabolic markers including inflammation, IR and secretion in women of normal pregnancy and with GDM. Our results suggested that individual FFAs have unique impacts on inflammation and glucose metabolism by mid-pregnancy.

In summary, our results found that individual FFAs are differentially associated with IR/secretion and inflammatory biomarkers. The findings suggest that specific FFAs play different roles in the regulation of glucose homeostasis in normal pregnancy and in women prior to the diagnosis of GDM. The effects are either direct or indirect depending on the modulating inflammatory response. Thus, the recognition and modification (possibly by dietary essential FA intake) of circulating FFA composition, especially the functional long-chain FFAs, is important for reducing the risk of GDM and other pregnancy complications related to subclinical inflammation and IR.

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takes responsibility for the integrity of the data and the accuracy of the data analysis.

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REFERENCES

- Homko CJ, Cheung P, Boden G. Effects of free fatty acids on glucose uptake and utilization in healthy women. *Diabetes* 2003;52:487–91.
- Sivan E, Homko CJ, Chen X, et al. Effect of insulin on fat metabolism during and after normal pregnancy. *Diabetes* 1999;48:834–8.
- Homko C, Sivan E, Chen X, et al. Insulin secretion during and after pregnancy in patients with gestational diabetes mellitus. *J Clin Endocrinol Metab* 2001;86:568–73.
- Barbour LA, McCurdy CE, Hernandez TL, et al. Cellular mechanisms for insulin resistance in normal pregnancy and gestational diabetes. *Diabetes Care* 2007;30(Suppl. 2):S112–9.
- Catalano PM, Huston L, Amini SB, et al. Longitudinal changes in glucose metabolism during pregnancy in obese women with normal glucose tolerance and gestational diabetes mellitus. *Am J Obstet Gynecol* 1999;180:903–16.
- McLachlan KA, Boston R, Alford FP. Impaired non-esterified fatty acid suppression to intravenous glucose during late pregnancy persists postpartum in gestational diabetes: a dominant role for decreased insulin secretion rather than insulin resistance. *Diabetologia* 2005;48:1373–9.
- Boden G, Chen X. Effects of fatty acids and ketone bodies on basal insulin secretion in type 2 diabetes. *Diabetes* 1999;48:577–83.
- Sivan E, Homko CJ, Whittaker PG, et al. Free fatty acids and insulin resistance during pregnancy. *J Clin Endocrinol Metab* 1998;83:2338–42.
- Tripathy D, Mohanty P, Dhindsa S, et al. Elevation of free fatty acids induces inflammation and impairs vascular reactivity in healthy subjects. *Diabetes* 2003;52:2882–7.
- Santaren ID, Watkins SM, Liese AD, et al. Individual serum saturated fatty acids and markers of chronic subclinical inflammation: the insulin resistance Atherosclerosis Study. *J Lipid Res* 2017;58:2171–9.
- Teng KT, Chang CY, Chang LF, et al. Modulation of obesity-induced inflammation by dietary fats: mechanisms and clinical evidence. *Nutr J* 2014;13.
- Crocker I, Lawson N, Daniels I, et al. Significance of fatty acids in pregnancy-induced immunosuppression. *Clin Diagn Lab Immunol* 1999;6:587–93.
- Steinberg HO, Tarshoby M, Monestel R, et al. Elevated circulating free fatty acid levels impair endothelium-dependent vasodilation. *J Clin Invest* 1997;100:1230–9.
- Haghiac M, Yang XH, Presley L, et al. Dietary omega-3 fatty acid supplementation reduces inflammation in obese pregnant women: a randomized double-blind controlled clinical trial. *PLoS One* 2015;10:e0137309.
- DeCaterina R, Liao JK, Libby P. Fatty acid modulation of endothelial activation. *Am J Clin Nutr* 2000;71(1 Suppl):213S–23.
- Jung UJ, Torrejon C, Chang CL, et al. Fatty acids regulate endothelial lipase and inflammatory markers in macrophages and in mouse aorta: a role for PPARgamma. *Arterioscler Thromb Vasc Biol* 2012;32:2929–37.
- Webster BH. Income, earnings, and Poverty data from the 2005 American community survey (ACS-02), 2006. Available: <https://www.census.gov/library/publications/2006/acs/acs-02.html>
- Chen X, Scholl TO. Ethnic differences in C-peptide/insulin/glucose dynamics in young pregnant women. *J Clin Endocrinol Metab* 2002;87:4642–6.
- Hauth JC, Clifton RG, Roberts JM, et al. Human development Maternal-Fetal Medicine units, N. maternal insulin resistance and preeclampsia. *Am J Obstet Gynecol* 2011;204:327e321–6.
- Chen X, Scholl TO. Maternal biomarkers of endothelial dysfunction and preterm delivery. *PLoS One* 2014;9:e85716.
- Wei SQ, Fraser W, Luo ZC. Inflammatory cytokines and spontaneous preterm birth in asymptomatic women: a systematic review. *Obstet Gynecol* 2010;116:393–401.
- American Diabetes Association. Clinical practice recommendations. *Diabetes Care* 2000;23(Suppl 1):S1–116.
- Chen X, Scholl TO, Leskiw M, et al. Differences in maternal circulating fatty acid composition and dietary fat intake in women with gestational diabetes mellitus or mild gestational hyperglycemia. *Diabetes Care* 2010;33:2049–54.
- Cohen O, Epstein GS, Weisz B, et al. Longitudinal assessment of insulin sensitivity in pregnancy. Validation of the homeostasis model assessment. *Clin Endocrinol* 2006;64:640–4.
- Lemaitre RN, Yu C, Hoofnagle A, et al. Circulating sphingolipids, insulin, HOMA-IR, and HOMA-B: the Strong Heart Family study. *Diabetes* 2018;67:1663–72.
- Hinkle SN, Tsai MY, Rawal S, et al. HbA1c measured in the first trimester of pregnancy and the association with gestational diabetes. *Sci Rep* 2018;8.
- Alptekin H, Cizmecioglu A, Isik H, et al. Predicting gestational diabetes mellitus during the first trimester using anthropometric measurements and HOMA-IR. *J Endocrinol Invest* 2016;39:577–83.
- Ozgu-Erdinc AS, Yilmaz S, Yeral MI, et al. Prediction of gestational diabetes mellitus in the first trimester: comparison of C-reactive protein, fasting plasma glucose, insulin and insulin sensitivity indices. *J Matern Fetal Neonatal Med* 2015;28:1957–62.
- Forouhi NG, Koulman A, Sharp SJ, et al. Differences in the prospective association between individual plasma phospholipid saturated fatty acids and incident type 2 diabetes: the EPIC-InterAct case-cohort study. *Lancet Diabetes Endocrinol* 2014;2:810–8.
- Baer DJ, Judd JT, Clevidence BA, et al. Dietary fatty acids affect plasma markers of inflammation in healthy men fed controlled diets: a randomized crossover study. *Am J Clin Nutr* 2004;79:969–73.
- Allaire J, Couture P, Leclerc M, et al. A randomized, crossover, head-to-head comparison of eicosapentaenoic acid and docosahexaenoic acid supplementation to reduce inflammation markers in men and women: the comparing EPA to DHA (compared) study. *Am J Clin Nutr* 2016;104:280–7.
- Fritsche KL. The science of fatty acids and inflammation. *Adv Nutr* 2015;6:293S–301.
- Chen X, O. Scholl T, Peter Stein T. Hypoadiponectinemia: association with risk of varying degrees of gestational hyperglycemia and with maternal ethnicity. *J Diabetes Mellitus* 2012;02:196–202.
- Lavoie F, Frisch F, Brassard P, et al. Relationship between total and high molecular weight adiponectin levels and plasma nonesterified fatty acid tolerance during enhanced intravascular triacylglycerol lipolysis in men. *J Clin Endocrinol Metab* 2009;94:998–1004.
- Fernandez-Real JM, Vendrell J, Ricart W. Circulating adiponectin and plasma fatty acid profile. *Clin Chem* 2005;51:603–9.
- Bernstein EL, Koutkia P, Ljungquist K, et al. Acute regulation of adiponectin by free fatty acids. *Metabolism* 2004;53:790–3.
- Arab L. Biomarkers of fat and fatty acid intake. *J Nutr* 2003;133:925S–32.
- Crowe FL, Skeaff CM, Green TJ, & Gray AR. Serum fatty acids as biomarkers of fat intake predict serum cholesterol concentrations in a population-based survey of New Zealand adolescents and adults. *Am J Clin Nutr* 2006;83:887–94.
- Laguzzi F, Alsharari Z, Riserus U, et al. Cross-sectional relationships between dietary fat intake and serum cholesterol fatty acids in a Swedish cohort of 60-year-old men and women. *J Hum Nutr Diet* 2016;29:325–37.
- Raatz SK, Bibus D, Thomas W, et al. Total fat intake modifies plasma fatty acid composition in humans. *J Nutr* 2001;131:231–4.
- Coustan DR, Lowe LP, Metzger BE, et al. International Association of D, & Pregnancy Study, G. The Hyperglycemia and Adverse Pregnancy Outcome (HAPO) study: paving the way for new diagnostic criteria for gestational diabetes mellitus. *Am J Obstet Gynecol* 2010;202:654.e651–6.
- d'Emden MC. Reassessment of the new diagnostic thresholds for gestational diabetes mellitus: an opportunity for improvement. *Med J Aust* 2014;201:209–11.